

# High cycling cadence reduces carbohydrate oxidation at given low intensity metabolic rate

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**ABSTRACT:** Cycling cadence (RPM)-related differences in blood lactate concentration (BLC) increase with increasing exercise intensity, whilst corresponding divergences in oxygen uptake ( $\dot{V}O_2$ ) and carbon dioxide production ( $\dot{V}CO_2$ ) decrease. Aim of the present study was to test whether a higher RPM reduces the fraction (%) of the  $\dot{V}O_2$  used for carbohydrate oxidation (relCHO) at a given BLC. Eight males ( $23.9 \pm 1.6$  yrs;  $177 \pm 3$  cm;  $70.3 \pm 3.4$  kg) performed incremental load tests at 50 and 100 RPM. BLC,  $\dot{V}O_2$  and  $\dot{V}CO_2$  were measured. At respiratory exchange ratios (RER) < 1, relCHO were calculated and the constant determining 50 % relCHO ( $k_{CHO}$ ) was approximated as a function of the BLC. At submaximal workload  $\dot{V}O_2$ ,  $\dot{V}CO_2$ , and relCHO were lower (all  $p < 0.002$ ;  $\eta^2 > 0.209$ ) at 50 than at 100 RPM. No differences were observed in  $\dot{V}O_{2peak}$  ( $3.96 \pm 0.22$  vs.  $4.00 \pm 0.25$  l·min<sup>-1</sup>) and RER<sub>peak</sub> ( $1.18 \pm 0.02$  vs.  $1.15 \pm 0.02$ ). BLC was lower ( $p < 0.001$ ;  $\eta^2 = 0.680$ ) at 50 than at 100 RPM irrespective of cycling intensity. At 50 RPM,  $k_{CHO}$  ( $4.2 \pm 1.4$  (mmol·l<sup>-1</sup>)<sup>3</sup>) was lower ( $p = 0.043$ ;  $\eta^2 = 0.466$ ) than at 100 RPM ( $5.9 \pm 1.9$  (mmol·l<sup>-1</sup>)<sup>3</sup>). This difference in  $k_{CHO}$  reflects a reduced CHO oxidation at a given BLC at 100 than at 50 RPM. At a low exercise intensity, a higher cycling cadence can substantially reduce the reliance on CHO at a given metabolic rate and/or BLC.

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## INTRODUCTION

Cycling cadence alterations are known to influence oxygen uptake ( $\dot{V}O_2$ ), carbon dioxide production ( $\dot{V}CO_2$ ), respiratory exchange ratio (RER) and levels of blood lactate concentration (BLC) over a wide range of exercise intensities [1,2,3]. BLC differences between varying cadences increase with exercise intensity. The differences in  $\dot{V}O_2$  and  $\dot{V}CO_2$  converge as exercise intensity increases with small or no differences at peak power ( $P_{peak}$ ) [3,4]. At cycling cadences of 90 to 100 revolutions per minute (RPM) this resulted in lower performances at the lactate threshold defined as an identifiable non-linear BLC-increase with increasing workload [4], at the second lactate deflection point [5], or at 2 and 4 mmol·l<sup>-1</sup> [6] compared to cadences of 40 to 60 RPM. However, top class professional road cyclists prefer cadences above 90 RPM during racing, testing in the laboratory and training [7-9] and at exercise intensities above 85 % of the maximum oxygen uptake they showed lower BLC,  $\dot{V}O_2$  and root-mean square EMG values at cadences at 100 RPM than at 60 RPM [8]. Prolonged constant load tests seem to show equivocal BLC, power output and intensity results at the transition from the heavy to the severe intensity domain as indicated by maximal lactate steady state results at different cadences [10-12]. While there is no differ-

ence in the critical power- $\dot{V}O_2$  relationship there is a lower critical power at a cycling cadence of 100 RPM compared with 60 RPM [13]. Whether the above partly equivocal cadence-effects on  $\dot{V}O_2$ ,  $\dot{V}CO_2$ , RER, BLC and sustainable mechanical power at prolonged exercise reflect also changes in the reliance on carbohydrates (CHO) as substrate of aerobic metabolism at a given exercise intensity related to  $P_{peak}$  ( $Int_p$ ) and also as fraction of  $\dot{V}O_{2peak}$  ( $Int_{\dot{V}O_2}$ ), is not known.

Aerobic CHO oxidation is regulated by the pyruvate dehydrogenase (PDH). Activators of the PDH are pyruvate, CoA and NAD<sup>+</sup>. Allosteric cofactors include Mg<sup>2+</sup>, Ca<sup>2+</sup> and Mn<sup>2+</sup> [14,15]. Substrate availability in terms of pyruvate and lactate is possibly one of the most potent effectors of PDH-activation, as food deprivation is known to shift metabolism to increased fat utilization. At a given metabolic rate, this results in a conservation of endogenous carbohydrates [16-21]. The corresponding decrease in muscular PDH-activation counts as a key regulator of this carbohydrate conserving effect. One identified factor of this muscle fibre-specific effect is the muscle fibre-specific PDH phosphatase profile [20].

Cycling cadence-dependent differences in cardio-respiratory and metabolic acute responses have been linked with muscle fibre acti-

vation. Higher fast twitch muscle fibre (FTF) recruitment is associated with higher cadences particularly at relative low exercise intensities [22-26]. Compared to the slow twitch fibre (STF) enzymatic profile, FTF have a higher content of anaerobic glycolytic enzymes combined with lower aerobic mitochondrial protein content [27-29]. These fibre-specific enzymatic properties result in lower PDH-activity combined with higher muscle lactate concentration in exercising muscles with approximately 70 % FTF compared with musculature of approximately 50 % FTF [28]. This higher PDH-activity in the more aerobic muscle has been related to the higher PDH-content rather than to a higher activation of the PDH [28] which in combination with a more marked dephosphorylation of the PDH-E1 $\alpha$  site 1 suggests that in FTF the PDH is less sensitive to the availability of pyruvate.

BLC,  $\dot{V}O_2$  and  $\dot{V}CO_2$  measurements are routine exercise testing measures. RER calculations and estimates of absolute and relative rates of fat and carbohydrate (relCHO) oxidation are often related to  $\dot{V}O_2$  and/or exercise intensity [30-34]. Compared to the concept of relating CHO-management to exercise intensity, the interpretation of relCHO as a function of the BLC and thus substrate availability is a change of paradigm. The bi-directional dynamic equilibrium between pyruvate and lactate is very much on the side of lactate. Thus the BLC serves as an indicator of the substrate activation of the whole body relCHO, which functionally reflects the mix of the activation of the PDH-complex of all aerobic tissues [10,35-37]. This approach was already used to address potential maturation-related differences in the reliance on CHO during exercise. It offered an explanation as to why under aerobic exercise conditions the reliance on CHO and the BLC are independent of maturation, whilst BLC but not relCHO is higher at high intensity exercise in more mature subjects [36].

Therefore the present study investigated the within subject effect of cycling cadence on the interrelationship between relCHO and BLC. The experiment was based firstly, on the frequently observed increase in cadence-related BLC-differences and the decreases in corresponding differences of  $\dot{V}O_2$  and  $\dot{V}CO_2$  at increasing exercise intensity [3,4]; secondly, the concept that the above observations reflect higher FTF-recruitment at higher cadences [24,26]; and thirdly, on data suggesting that in FTF the PDH is less sensitive to the availability of pyruvate as substrate of mitochondrial metabolism [28]. The above background led to the hypothesis that compared with lower cadences a higher cycling cadence causes a higher metabolic rate, a higher BLC and a higher relCHO at a given mechanical power but a lower relCHO at given BLC-levels.

## MATERIALS AND METHODS

**Subjects.** Eight healthy males (Tab. 1) volunteered and gave written informed consent to participate in this study, which was approved by the university's ethics committee. All experiments were performed in accordance with the ethical standards of the Helsinki declaration. The subjects were instructed to avoid any strenuous exercise or alcohol consumption, and consume the same diet for the 24 hours

preceding a test, and to arrive in a fully hydrated state without consuming any heavy meals for at least 2 hours prior to the test. Subjects were familiarized with all testing procedures prior to the testing day. All tests were performed at a similar time of day.

### Exercise testing

All subjects arrived at the laboratory approximately one hour before testing. Test preparation included a short interview about exercise, sleep, food intake and drinking during the previous 24 hours to exclude issues likely to affect the experimental results. Subjects completed two separate incremental load cycling tests at 50 and 100 RPM on an electro-magnetically braked cycle Ergometer (Lode Excalibur Sport, The Netherlands) in a counter balanced random order. The tests were completed within a period of two weeks and under similar environmental conditions ( $19.0 \pm 0.3$  °C,  $1022 \pm 4$  mmHg and  $55 \pm 3\%$  humidity). Seat and handle bar positions were recorded for the first test and reproduced for the subsequent test. Each test started with a mechanical power of  $1.0 \text{ W} \cdot \text{kg}^{-1}$  body mass, which was increased by  $0.5 \text{ W} \cdot \text{kg}^{-1}$  after every second minute. Subjects cycled until volitional exhaustion, defined as the inability to maintain the required cycling cadence for longer than 15 seconds despite verbal encouragement.

### Measurements

$\dot{V}O_2$  and  $\dot{V}CO_2$  were continuously measured breath-by-breath throughout the incremental power test (Oxycon Gamma, Mijnhard, The Netherlands). Flow sensor and gas analyzers were calibrated using a 3 l syringe and gases of known concentration prior to each test.

**TABLE I.** Anthropometric data, peak performance and corresponding physiological acute response

	50 RPM (mean $\pm$ SE)	100 RPM (mean $\pm$ SE)	Sig.	$\eta^2$
Age (years)	23.9 $\pm$ 1.6			
Height (cm)	177 $\pm$ 3			
Body mass (kg)	68.3 $\pm$ 3.5			
$P_{\text{peak}}$ (W)	290 $\pm$ 18	286 $\pm$ 19		
rel $P_{\text{peak}}$ ( $\text{W} \cdot \text{kg}^{-1}$ )	4.3 $\pm$ 0.2	4.2 $\pm$ 0.2		
$\dot{V}O_{2\text{peak}}$ ( $\text{l} \cdot \text{min}^{-1}$ )	3.96 $\pm$ 0.22	4.00 $\pm$ 0.25		
rel $\dot{V}O_{2\text{peak}}$ ( $\text{ml} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ )	58.2 $\pm$ 1.7	58.7 $\pm$ 2.1		
BLC $_{\text{peak}}$ ( $\text{mmol} \cdot \text{l}^{-1}$ )	10.0 $\pm$ 0.5	11.8 $\pm$ 0.7	p=0.007	0.669
RER $_{\text{peak}}$	1.18 $\pm$ 0.02	1.15 $\pm$ 0.02		

Note: Peak power ( $P_{\text{peak}}$ ), Peak power related to body mass (rel $P_{\text{peak}}$ ), Peak oxygen uptake ( $\dot{V}O_{2\text{peak}}$ ), Peak oxygen uptake related to body mass (rel $\dot{V}O_{2\text{peak}}$ ), highest blood lactate concentration measured during the post-exercise period (BLC $_{\text{peak}}$ ), respiratory exchange ratio at  $P_{\text{peak}}$  (RER $_{\text{peak}}$ )

The breath-by-breath oxygen uptake data were reduced to stationary averages of the final 30 s of each stage.

Capillary blood samples (20  $\mu$ l) for BLC analysis were collected from the hyperaemic (Finalgon forte®, Thomae) ear lobe during the final 15 s of each stage and every minute after test termination up to the fifth minute. All BLC samples were immediately analyzed utilizing the enzymatic amperometric method (Ebio Plus, Eppendorf).

*Data processing and statistics*

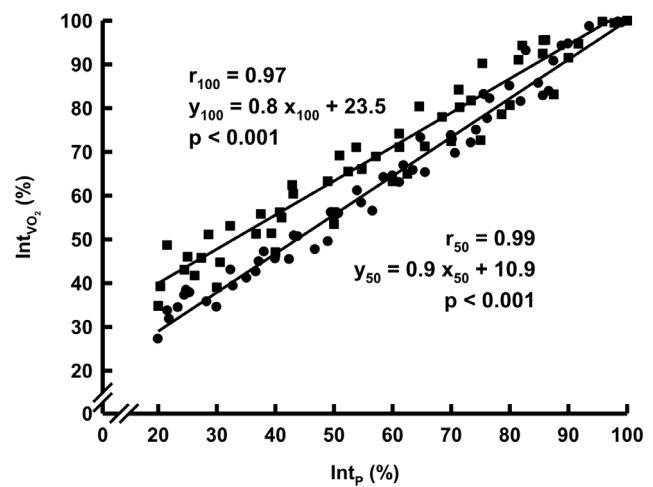
Peak power ( $P_{peak}$ ) was equivalent to the power at the final stage if a test was terminated after completion of the full stage. If the test was terminated before 2 min had been completed,  $P_{peak}$  was calculated as:  $P_{peak}$  (W) = power of previous stage (W) plus power increment (W) times duration of exercise at the final stage (s) divided by 120 s.  $\dot{V}O_{2peak}$  ( $ml \cdot min^{-1}$ ) was defined as the highest  $\dot{V}O_2$  averaged throughout a 30 s time segment of the final minute of the test.  $BLC_{peak}$  ( $mmol \cdot l^{-1}$ ) was determined as the highest BLC measured during the post-test period.  $Int_p$  and  $Int_{\dot{V}O_2}$  were calculated as percentage of  $P_{peak}$  and  $\dot{V}O_{2peak}$ , respectively. CHO was calculated from  $\dot{V}O_2$  ( $ml \cdot min^{-1}$ ) and  $\dot{V}CO_2$  ( $ml \cdot min^{-1}$ ) measured at each stage via indirect calorimetry [38]. The relCHO was calculated as the fraction (%) of the  $\dot{V}O_2$  used for CHO with 100 % reflecting a respiratory exchange ratio (RER)  $\geq 1.0$ . As a first approximation relCHO was individually plotted as a function of the BLC ( $relCHO = 100\% / (1 + k_{CHO}/BLC^n)$ ) [10,35-37].  $k_{CHO}$  is defined as the constant determining relCHO of 50 %, and n is set 3.

Descriptive results are presented as mean  $\pm$  SE. Respiratory and BLC data at  $P_{peak}$  and at all stages below  $P_{peak}$ , and relCHO of each stage with an RER < 1.0 were tested for normal distribution using Kolmogorov-Smirnov test. Peak data of 50 RPM and 100 RPM tests were compared using paired t-test. A cadence-by-power ANOVA analysis was used to analyze all sub-peak data measured with 'cadence' as within and 'power' as between factor. Significant interactions and main effects were further analyzed using t-tests and paired samples t-tests as appropriate. Based on cadence and power effects on  $\dot{V}O_2$  and BLC observed previously [39], assuming normal distribution an a priori power calculation revealed the necessity of a sample size of n = 8 to achieve a power of 80 % at a significance level of  $p \leq 0.05$ . Effect sizes in the form of eta<sup>2</sup> ( $\eta^2$ ) were calculated.

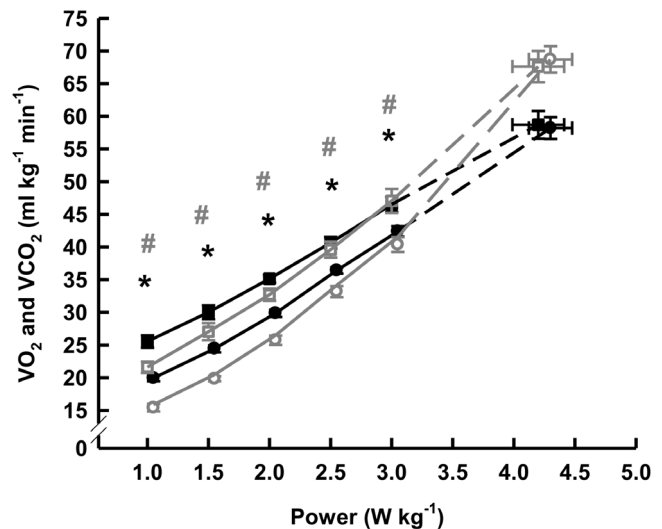
**RESULTS**

$P_{peak}$ , peak respiratory data and  $BLC_{peak}$ , and all corresponding data at power stages below  $P_{peak}$  were normal distributed.  $P_{peak}$ ,  $\dot{V}O_{2peak}$  and  $RER_{peak}$  were not different between cycling cadences (Tab.1). There were main effects of cadence and power in  $\dot{V}O_2$  ( $p < 0.001$ ;  $\eta^2 = 0.528$  and  $p < 0.001$ ;  $\eta^2 = 0.874$ ),  $\dot{V}CO_2$  ( $p < 0.001$ ;  $\eta^2 = 0.472$  and  $p < 0.001$ ;  $\eta^2 = 0.907$ ), BLC ( $p < 0.001$ ;  $\eta^2 = 0.600$  and  $p < 0.001$ ;  $\eta^2 = 0.640$ ) and relCHO ( $p = 0.002$ ;  $\eta^2 = 0.209$  and  $p < 0.001$ ;  $\eta^2 = 0.785$ ) with interaction of cadence and power in BLC ( $p = 0.042$ ;  $\eta^2 = 0.280$ ) and relCHO ( $p = 0.043$ ;  $\eta^2 = 0.278$ ).

At both cadences, the  $\dot{V}O_2$  and the  $\dot{V}CO_2$  increased with each increase in workload ( $p < 0.01$ ). A corresponding increase in the BLC was significant above a workload of  $2.5 W \cdot kg^{-1}$  whilst relCHO increased from 1.5 to  $2.5 W \cdot kg^{-1}$  irrespective of cadence. At both cadences,  $Int_p$  and  $Int_{\dot{V}O_2}$  were highly correlated, however, inclination of the regression line was steeper at 50 RPM than at 100 RPM resulting in higher  $Int_{\dot{V}O_2}$  at given  $Int_p$  at 100 RPM than at 50 RPM which diminished towards  $P_{peak}$  (Fig. 1). Consequently, at a given sub-maximal workload  $\dot{V}O_2$  but also  $\dot{V}CO_2$  were lower at 50 RPM than at 100 RPM (Fig. 2).



**FIG. 1.** Exercise intensity related to  $\dot{V}O_{2peak}$  ( $Int_{\dot{V}O_2}$ ) as a function of exercise intensity related to  $P_{peak}$  ( $Int_p$ ) at 50 RPM (●) and at 100 RPM (■).



**FIG. 2.**  $\dot{V}O_2$  (black) and  $\dot{V}CO_2$  (grey) at 50 RPM (● and ○) and at 100 RPM (■ and □); \* =  $\dot{V}O_2$  difference between 50 and 100 RPM; mean  $\pm$  SE; # =  $\dot{V}CO_2$  difference between 50 and 100 RPM; all  $p < 0.05$ .

The BLC was lower ( $p < 0.001$ ) at 50 RPM than at 100 RPM irrespective of workload (Fig. 3). A corresponding effect on relCHO was seen up to a workload of  $2.0 \text{ W} \cdot \text{kg}^{-1}$  (Fig. 3), which was equivalent to BLC levels of  $1.8 \pm 0.2 \text{ mmol} \cdot \text{l}^{-1}$  vs.  $2.7 \pm 0.3 \text{ mmol} \cdot \text{l}^{-1}$  ( $p = 0.003$ ;  $\eta^2 = 0.745$ ), and  $\text{Int}_{\text{VO}_2}$  of  $51.6 \pm 1.4 \%$  vs.  $60.5 \pm 2.5 \%$  ( $p < 0.01$ ), whilst  $\text{Int}_{\text{p}}$  of  $47.5 \pm 1.8 \%$  vs.  $48.4 \pm 2.3 \%$  (n.s.) were not different at 50 and 100 RPM. At the individually highest workload ( $2.9 \pm 0.2 \text{ W} \cdot \text{kg}^{-1}$  vs.  $2.8 \pm 0.3 \text{ W} \cdot \text{kg}^{-1}$ ; n.s.) with an  $\text{RER} < 1.0$  ( $0.97 \pm 0.01$  vs.  $0.99 \pm 0.01$ ; n.s.), the BLC was lower ( $2.9 \pm 0.3 \text{ mmol} \cdot \text{l}^{-1}$  vs.  $4.0 \pm 0.3 \text{ mmol} \cdot \text{l}^{-1}$ ;  $p = 0.009$ ;  $\eta^2 = 0.651$ ) at 50 RPM than at 100 RPM but not  $\text{Int}_{\text{p}}$  ( $68.6 \pm 2.4 \%$

vs.  $64.5 \pm 3.4 \%$ ; n.s.) and  $\text{Int}_{\text{VO}_2}$  ( $71.4 \pm 3.0 \%$  vs.  $74.0 \pm 3.0 \%$ ; n.s.) or relCHO ( $89.5 \pm 1.9 \%$  vs.  $95.1 \pm 1.2 \%$ ; n.s.).

At 50 RPM  $k_{\text{CHO}}$  ( $k_{\text{CHO}50}$ :  $4.2 \pm 1.4 \text{ (mmol l}^{-1}\text{)}^3$ ) was lower ( $p = 0.043$ ;  $\eta^2 = 0.466$ ) than the  $k_{\text{CHO}}$  at 100 RPM ( $k_{\text{CHO}100}$ :  $5.9 \pm 1.9 \text{ (mmol l}^{-1}\text{)}^3$ ). The approximation model explained  $90 \pm 3$  and  $91 \pm 4 \%$  of the variance of the relCHO at 50 RPM and 100 RPM, respectively (both  $p < 0.001$ , Fig. 4).  $k_{\text{CHO}50}$  and  $k_{\text{CHO}100}$  were independent of  $P_{\text{peak}}$ ,  $\dot{\text{V}}\text{O}_{2\text{peak}}$ ,  $\text{RER}_{\text{peak}}$  or  $\text{BLC}_{\text{peak}}$  and also the difference between  $k_{\text{CHO}50}$  and  $k_{\text{CHO}100}$  was independent of  $P_{\text{peak}}$  and  $\dot{\text{V}}\text{O}_{2\text{peak}}$ .

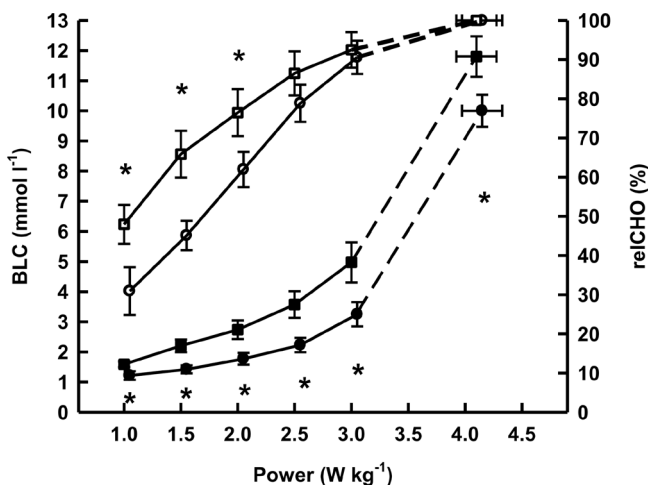
**DISCUSSION**

The main new finding of the present study was that at a higher cycling cadence the relCHO was reduced at given BLC-levels as indicated by a higher  $k_{\text{CHO}}$  (Fig. 4). This cadence-related CHO-preserving effect was apparent below approximately 50 %  $\text{Int}_{\text{p}}$  only.

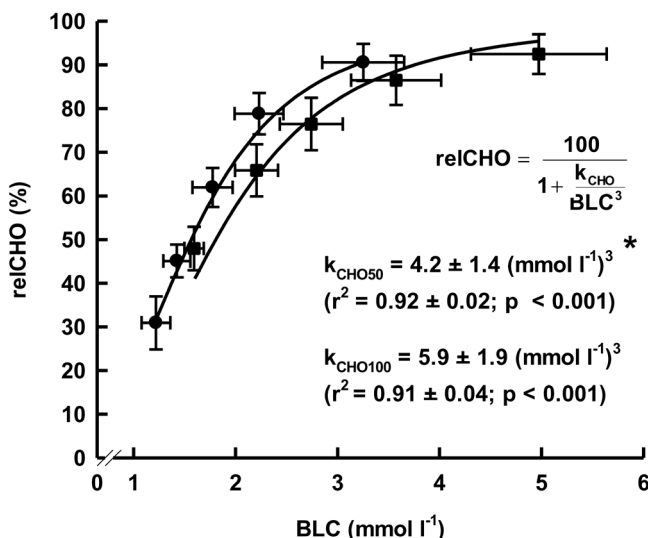
The present findings are consistent with previous observations of higher  $\dot{\text{V}}\text{O}_2$ ,  $\dot{\text{V}}\text{CO}_2$ ,  $\text{RER}$  and BLC values at higher cadences over a wide range of given sub-maximal exercise intensities [1-3]. They confirm that BLC-differences between different cadences increase with exercise intensity, whereas the differences in  $\dot{\text{V}}\text{O}_2$  and  $\dot{\text{V}}\text{CO}_2$  converge as exercise intensity increases with small or no differences at  $P_{\text{peak}}$  [3,4].

$\dot{\text{V}}\text{O}_2$ ,  $\dot{\text{V}}\text{CO}_2$ ,  $\text{RER}$  and BLC values do not directly reflect metabolism at muscle fiber level. Respiratory measures represent metabolism of all aerobic tissues and the BLC indicates corresponding substrate availability in its dilution space. In the third decade of life, healthy males have an average relative skeletal muscle mass related to the total body mass of about 40 % [40]. In comparison to the skeletal muscle other tissues with lactate release and oxidation like heart muscle, liver, kidney, brain [41,42] are comparably small or less well perfused during exercise than skeletal muscle. The exercise-induced and cadence-related increase in the metabolic rate at an  $\text{RER} < 1$  was approximately up to four to eight fold the resting metabolic rate at 50 RPM and five to nine fold at 100 RPM (Fig. 2). The corresponding BLC-levels were up to approximately three and four times the resting value, respectively. Consequently, skeletal muscle can be seen as the dominant factor of the cycling-induced metabolic response seen in the present study.

The observed within subject differences in the metabolic response at given workload between cycling at 50 vs. 100 RPM below approximately 50 %  $\text{Int}_{\text{p}}$  are indicative for increased muscle fibre activation at the higher cadence, e.g. a higher fast twitch muscle fibre (FTF) recruitment associated with higher cadences [22-26]. This effect should decrease, disappear or even reverse with increasing exercise intensity [26] as seen in subjects with varied athletic background [24,43] and professional road cyclists [8] cycling at and above 85 %  $\dot{\text{V}}\text{O}_{2\text{peak}}$ . Other factors which may also affect fibre recruitment during cycling at given workload including muscle strength [44], cycling skills [45], saddle position [46,47], pedal design [48] and test duration [23] have been minimized or excluded through the within subject design and standardization in the present experiment.



**FIG. 3.** Blood lactate concentration (BLC) and relative carbohydrate oxidation (relCHO) at 50 RPM (● and ○) and at 100 RPM (■ and □); mean ± SE; \* = difference between 50 and 100 RPM (all  $p < 0.05$ ).



**FIG. 4.** Relative carbohydrate oxidation (relCHO) related to the blood lactate concentration (BLC) at 50 RPM (●) and 100 RPM (■); mean ± SE; \* = difference between 50 und 100 RPM ( $p < 0.05$ ).

BLC and  $\text{Int}_{\dot{V}O_2}$  are common measures of exercise intensity of recreational and high performance training [49]. The newly described difference in the BLC-relCHO-relationship indicates, that at a given  $\dot{V}O_2$  and/or BLC higher cadences reduce the reliance on CHO as a metabolic substrate of aerobic energy (Fig. 4). Comparable effects on BLC and RER, and a lower use of CHO as metabolic substrate of aerobic energy at given workload have been described at incremental and constant workload exercise in glycogen depletion cycling experiments [50,51]. Such a carbohydrate conserving effect was linked with a decreased muscular PDH-activation [16-21,28]. The localization of glycogen, the pattern of depletion of the muscle cell and carbohydrate availability lead to a fibre type-specific compartmentalization of glycogen metabolism [52,53]. Also muscle fibre-specific PDH phosphatase profiles have been described [20]. The higher PDH-activity in the more aerobic muscle was related to the higher PDH-content [54]. Higher content of anaerobic glycolytic enzymes, lower aerobic mitochondrial protein content of FTF compared with STF [27,29,54], and higher muscle lactate concentration combined with lower PDH-activity in FTF suggest that the PDH of FTF is less sensitive to the availability of pyruvate than STF [54].

The higher BLC and  $k_{\text{CHO}}$  are therefore consistent with suggestions that cycling cadence-dependent differences in cardio-respiratory and metabolic acute responses reflect higher FTF-recruitment at higher cadences [22-26]. Consequently, at a given metabolic rate the higher BLC, as observed at 100 RPM compared with 50 RPM, does not necessarily reflect a higher glycolytic rate but a reduced sensitivity to the availability of substrate requiring a higher equilibrium concentration of pyruvate and lactate as a substrate of the aerobic re-phosphorylation of the whole body aerobic tissue with a cadence-related higher fraction of activated FTF.

Racing road cyclists prefer cadences above 90 RPM during racing, testing in the laboratory and training [7-9]. Depending on seasonal variations, top athletes competing in long distance events train 70 to 90% of their endurance training at intensities corresponding to BLCs below  $2 \text{ mmol} \cdot \text{l}^{-1}$  [49,55]. The present cadence-related CHO-preserving effect of a higher  $k_{\text{CHO}}$  at a higher cadence is most relevant at BLC-levels up to  $2.0 \text{ mmol} \cdot \text{l}^{-1}$  (Fig. 3 and 4) comparable to approximately 50 % of  $\dot{V}O_{2\text{peak}}$  (Fig. 2) where relCHO is approximately 65 % (equivalent to  $25 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ ) at 50 RPM and 58 % (equivalent to  $22 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ ) at 100 RPM. This CHO-saving effect of

approximately 12 % disappears at an  $\text{Int}_{\dot{V}O_2}$  of approximately 70 %, which is roughly comparable to an  $\text{Int}_p$  of 60 % with corresponding relCHO of 82 and 86 % (equivalent to  $46 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$  vs.  $44 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$  CHO oxidation) at 50 and 100 RPM, respectively, above which relCHO approaches saturation. Many endurance athletes monitor their exercise intensity via blood lactate measurements adjusted to exercise intensity domains expressed as a fraction of the peak oxygen uptake rather than mechanical power [49,55].  $k_{\text{CHO}50}$  and  $k_{\text{CHO}100}$  and also the cadence-effect on  $k_{\text{CHO}}$  were independent of both  $P_{\text{peak}}$  and  $\dot{V}O_{2\text{peak}}$  suggesting that in top cyclists athletes with an up to 40 % higher  $\dot{V}O_{2\text{peak}}$  than the present subjects combined with up to 30 hours of training per week such a cadence related CHO preserving effect may become a protective factor against glycogen depletion.

## CONCLUSIONS

Frequently observed higher  $\dot{V}O_2$ ,  $\dot{V}CO_2$  and BLC values at higher cycling cadences over a wide range of exercise intensities suitable for high training volumes are combined with a reduced relCHO at given BLC-values as indicated by a higher  $k_{\text{CHO}}$ . These findings are consistent with suggestions that at such exercise intensities cycling cadence-dependent differences in cardio-respiratory and metabolic acute responses reflect higher FTF recruitment at higher cadences [1-3], and that the PDH of FTF is less sensitive to the availability of pyruvate than that of STF [54]. The present results also suggest that in spite of a higher  $\dot{V}O_2$ , a higher BLC and a higher CHO oxidation at a given mechanical power compared to a low cycling cadence, a higher cadence can substantially reduce the reliance on CHO at a given low exercise intensity as indicated by a particular  $\dot{V}O_2$  and/or BLC.

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**Conflict of interests:** The authors declare that there exist no conflicts of interest.

## REFERENCES

1. Chavarran J, Calbet JAL. Cycling efficiency and pedaling frequency in road cyclists. *Eur J Appl Physiol* 1999;80:555-563.
2. Gaesser GA, Brooks GA. Muscular efficiency during steady rate: effects of speed and work rate. *J Appl Physiol* 1975;38:1132-1139.
3. Zoladz JA, Rademaker AC, Sargeant AJ. Human muscle power generating capability during cycling at different pedalling rates. *Exp Physiol* 2000;85:117-24.
4. Hughes EF, Turner SC, Brooks GA. Effect of glycogen depletion and pedalling speed on anaerobic threshold. *J Appl Physiol* 1982;52:1598-1607.
5. Woolford SM, Withers RT, Craig NP, Bourdon PC, Stanef T, McKenzie I. Effect of pedal cadence on the accumulated oxygen deficit, maximal aerobic power and blood lactate transition thresholds of high-performance junior endurance cyclists. *Eur J Appl Physiol Occup Physiol* 1999;80:285-91.
6. Buchanan M, Weltman A. Effects of pedal frequency on  $\dot{V}O_2$  and work output at lactate threshold (LT), fixed blood lactate concentrations of 2 mM and 4 mM, and max in competitive cyclists. *Int J Sports Med* 1985;6:163-168.
7. Gueli D, Shephard RJ. Pedal frequency in bicycle ergometry. *Can J Appl Sport Sci* 1976;1:137-141.

8. Lucia A, San Juan AF, Montilla M, CaNete S, Santalla A, Earnest C, Pérez M. In professional road cyclists, low pedaling cadences are less efficient. *Med Sci Sports Exerc* 2004;36:1048-54.
9. Palmer GS, Noakes TD, Hawley JA. Metabolic and performance responses to constant-load vs variable-intensity exercise in trained cyclists. *J Appl Physiol* 1999;87:1186-96.
10. Beneke R. Maximal lactate steady state concentration (MLSS): experimental and modelling approaches. *Eur J Appl Physiol* 2003a;88:361-369.
11. Beneke R, von Duvillard SP. Determination of maximal lactate steady state response in selected sports events. *Med Sci Sports Exerc* 1996;28:241-246.
12. Denadai BS, Ruas VD, Figueira TR. Maximal lactate steady state concentration independent of pedal cadence in active individuals. *Eur J Appl Physiol* 2006;96:477-480.
13. Barker T, Poole DC, Noble ML, Barstow TJ. 2006. Human critical power-oxygen uptake relationship at different pedaling frequencies. *Exp Physiol* 2006;91:621-632.
14. Spriet LL, Heigenhauser GJ. Regulation of pyruvate dehydrogenase (PDH) activity in human skeletal muscle during exercise. *Exerc Sport Sci Rev* 2002;30:91-95.
15. Strumilo S. Short-term regulation of the mammalian pyruvate dehydrogenase complex. *Acta Biochim Pol* 2005;52:759-764.
16. Denyer GS, Lam D, Cooney GJ, Caterson ID. Effect of starvation and insulin in vivo on the activity of the pyruvate dehydrogenase complex in rat skeletal muscle. *FEBS Lett* 1989;250:464-468.
17. Fuller SJ, Randle PJ. Reversible phosphorylation of pyruvate dehydrogenase in rat skeletal-muscle mitochondria. Effects of starvation and diabetes. *Biochem J* 1984;219:635-646.
18. Hagg SA, Taylor SI, Ruderman NB. Glucose metabolism in perfused skeletal muscle. Pyruvate dehydrogenase activity in starvation, diabetes and exercise. *Biochem J* 1976;158:203-210.
19. Holness MJ, Liu YL, Sugden MC. Time courses of the responses of pyruvate dehydrogenase activities to short-term starvation in diaphragm and selected skeletal muscles of rat. *Biochem J* 1989;264:771-776.
20. LeBlanc PJ, Harris RA, Peters SJ. Skeletal muscle fiber type comparison of pyruvate dehydrogenase phosphatase activity and isoform expression in fed and food-deprived rats. *Am J Physiol Endocrinol Metab* 2006; 92:E571-E576.
21. Sugden MC, Kraus A, Harris RA, Holness MJ. Fibre-type specific modification of the activity and regulation of skeletal muscle pyruvate dehydrogenase kinase (PDK) by prolonged starvation and refeeding is associated with targeted regulation of PDK isoenzyme 4 expression. *Biochem J* 2000;346:651-657.
22. Dantas JL, Smirmaul BP, Altimari LR, Okano AH, Fontes EB, Camata TV, Moraes AC. The efficiency of pedaling and the muscular recruitment are improved with increase of the cadence in cyclists and non-cyclists. *Electromyogr Clin Neurophysiol* 2009;49(6-7):311-319.
23. Kounalakis SN, Geladas ND. Cardiovascular drift and cerebral and muscle tissue oxygenation during prolonged cycling at different pedalling cadences. *Appl Physiol Nutr Metab* 2012;37(3):407-17.
24. Macintosh BR, Neptune RR, Horton JF. Cadence, power, and muscle activation in cycle ergometry. *Med Sci Sports Exerc* 2000;32:1281-1287.
25. Sanderson DJ, Martin PE, Honeyman G, Keefer J. Gastrocnemius and soleus muscle length, velocity, and EMG responses to changes in pedalling cadence. *J Electromyogr Kinesiol* 2006;16(6):642-649.
26. Sargeant AJ. Human power output and muscle fatigue. *Int J Sports Med* 1994;15:116-121.
27. Essen-Gustavsson B, Henriksson J. Enzyme levels in pools of microdissected human muscle fibres of identified type. Adaptive response to exercise. *Acta Physiol Scand* 1984;120:505-515.
28. Kiilerich K, Gudmundsson M, Birk J, Lundby C, Taudorf S, Plomgaard P, Saltin B, Pedersen PA, Wojtaszewski JFP, Pilegaard H. Low muscle glycogen and elevated plasma free fatty acid modify but do not prevent exercise-induced PDH activation in human skeletal muscle. *Diabetes*. 2010;59:26-32.
29. Plomgaard P, Penkowa M, Leick L, Pedersen BK, Saltin B, Pilegaard H. The mRNA expression profile of metabolic genes relative to MHC isoform pattern in human skeletal muscles. *J Appl Physiol* 2006;101:817-825.
30. Martinez LR, Haymes EM. Substrate utilization during treadmill running in prepubertal girls and women. *Med Sci Sports Exerc* 1992;24:975-983.
31. Timmons BW, Bar-Or O, Riddell MC. Oxidation rate of exogenous carbohydrate during exercise is higher in boys than in men. *J Appl Physiol* 2003;94:278-284.
32. Timmons BW, Bar-Or O, Riddell MC. Influence of age and pubertal status on substrate utilization during exercise with and without carbohydrate intake in healthy boys. *Appl Physiol Nutr Metab* 2007a;32:416-425.
33. Timmons BW, Bar-Or O, Riddell MC. Energy substrate utilization during prolonged exercise with and without carbohydrate intake in preadolescent and adolescent girls. *J Appl Physiol* 2007b;103:995-1000.
34. Venables MC, Achten J, Jeukendrup AE. Determinants of fat oxidation during exercise in healthy men and women: a cross-sectional study. *J Appl Physiol* 2005;98:160-167.
35. Beneke R. Experiment and computer aided simulation complementary tools to understand exercise metabolism. *Biochem Soc Trans* 2003b;31:1263-1266.
36. Beneke R, Hütler M, Leithäuser RM. Carbohydrate and fat metabolism related to blood lactate in boys and male adolescents. *Eur J Appl Physiol* 2009;105:257-263. doi: 10.1007/s00421-008-0897-1.
37. Mader A, Heck H. A theory of the metabolic origin of „anaerobic threshold“. *Int J Sports Med* 1986;7(Suppl 1):45-65.
38. Péronnet F, Massicotte D. Table of nonprotein respiratory quotient: an update. *Can J Sport Sci* 1991;16:23-29.
39. Beneke R. *Laktat bei Dauerleistung. Habilitationsschrift. Berlin; 1999.*
40. Janssen I, Heymsfield SB, Wang ZM, Ross R. Skeletal muscle mass and distribution in men and women aged 18-88 yr. *J Appl Physiol* 2000;89:81-88.
41. Barros LF. Metabolic signaling by lactate in the brain. *Trends Neurosci* 2013;36(7):396-404.
42. Walsh ML, Banister EW. Possible mechanisms of the anaerobic threshold. A review. *Sports Med* 1988;5(5):269-302.
43. Ahlquist LE, Bassett DR Jr, Sufit R, Nagle FJ, Thomas DP. The effect of pedaling frequency on glycogen depletion rates in type I and type II quadriceps muscle fibers during submaximal cycling exercise. *Eur J Appl Physiol Occup Physiol*. 1992;65(4):360-364.
44. Bieuzen F, Lepers R, Vercauysen F, Hausswirth C, Brisswalter J. Muscle activation during cycling at different cadences: effect of maximal strength capacity. *J Electromyogr Kinesiol* 2007;17(6):731-738.
45. Chapman AR, Vicenzino B, Blanch P, Hodges PW. Patterns of leg muscle recruitment vary between novice and highly trained cyclists. *J Electromyogr Kinesiol* 2008;18(3):359-371.
46. Sanderson DJ, Amoroso AT. The influence of seat height on the mechanical function of the triceps surae muscles during steady-rate cycling. *J Electromyogr Kinesiol* 2009;19(6):e465-471.

47. Fonda B, Panjan A, Markovic G, Sarabon N. Adjusted saddle position counteracts the modified muscle activation patterns during uphill cycling. *J Electromyogr Kinesiol* 2011;21(5):854-860. doi: 10.1016/j.jelekin.2011.05.010.
48. Cruz CF, Bankoff AD. Electromyography in cycling: difference between clipless pedal and toe clip pedal. *Electromyogr. Clin Neurophysiol* 2001;41(4):247-52.
49. Beneke R, Leithäuser RM, Ochentel O. Blood Lactate Diagnostics in Exercise Testing and Training. *Int J Sports Physiol Perform* 2011;6:8-24.
50. Lima-Silva AE, De-Oliveira FR, Nakamura FY, Gevaerd MS. Effect of carbohydrate availability on time to exhaustion in exercise performed at two different intensities. *Braz J Med Biol Res* 2009;42:404-12.
51. Mikulski T, Ziemba A, Nazar K. Influence of body carbohydrate store modification on catecholamine and lactate responses to graded exercise in sedentary and physically active subjects. *J Physiol Pharmacol* 2008;59:603-16.
52. Nielsen J, Holmberg HC, Schrøder HD, Saltin B, Ørtenblad N. Human skeletal muscle glycogen utilization in exhaustive exercise: role of subcellular localization and fibre type. *J Physiol* 2011;589:2871-2885.
53. Ørtenblad N, Nielsen J, Saltin B, Holmberg HC. Role of glycogen availability in sarcoplasmic reticulum Ca<sup>2+</sup> kinetics in human skeletal muscle. *J Physiol* 2011;589:711-725. doi: 10.1113/jphysiol.2010.195982.
54. Kiilerich K, Birk JB, Damsgaard R, Wojtaszewski JP, Pilegaard H. Regulation of PDH in human arm and leg muscles at rest and during intense exercise. *Am J Physiol Endocrinol Metab* 2007;294:E36-E42.
55. Seiler S. What is best practice for training intensity and duration distribution in endurance athletes? *Int J Sports Physiol Perform* 2010;5:276-291.